

## ARTICLE

Received 6 Jun 2014 | Accepted 8 Jul 2014 | Published 6 Aug 2014

DOI: 10.1038/ncomms5619

OPEN

# Matrix softness regulates plasticity of tumour-repopulating cells via H3K9 demethylation and Sox2 expression

Youhua Tan<sup>1,2,\*</sup>, Arash Tajik<sup>2,\*</sup>, Junwei Chen<sup>1,\*</sup>, Qiong Jia<sup>1</sup>, Farhan Chowdhury<sup>3</sup>, Lili Wang<sup>1</sup>, Junjian Chen<sup>1</sup>, Shuang Zhang<sup>1</sup>, Ying Hong<sup>1</sup>, Haiying Yi<sup>1</sup>, Douglas C. Wu<sup>2</sup>, Yuejin Zhang<sup>1</sup>, Fuxiang Wei<sup>1</sup>, Yeh-Chuin Poh<sup>1,2</sup>, Jihye Seong<sup>2,4,†</sup>, Rishi Singh<sup>2</sup>, Li-Jung Lin<sup>4</sup>, Sultan Doğanay<sup>3,5</sup>, Yong Li<sup>6</sup>, Haibo Jia<sup>1</sup>, Taekjip Ha<sup>3,5</sup>, Yingxiao Wang<sup>7</sup>, Bo Huang<sup>6,8</sup> & Ning Wang<sup>1,2</sup>

Tumour-repopulating cells (TRCs) are a self-renewing, tumorigenic subpopulation of cancer cells critical in cancer progression. However, the underlying mechanisms of how TRCs maintain their self-renewing capability remain elusive. Here we show that relatively undifferentiated melanoma TRCs exhibit plasticity in Cdc42-mediated mechanical stiffening, histone 3 lysine residue 9 (H3K9) methylation, Sox2 expression and self-renewal capability. In contrast to differentiated melanoma cells, TRCs have a low level of H3K9 methylation that is unresponsive to matrix stiffness or applied forces. Silencing H3K9 methyltransferase G9a or SUV39h1 elevates the self-renewal capability of differentiated melanoma cells in a Sox2-dependent manner. Mechanistically, H3K9 methylation at the Sox2 promoter region inhibits Sox2 expression that is essential in maintaining self-renewal and tumorigenicity of TRCs both *in vitro* and *in vivo*. Taken together, our data suggest that 3D soft-fibrin-matrix-mediated cell softening, H3K9 demethylation and Sox2 gene expression are essential in regulating TRC self-renewal.

<sup>1</sup>Laboratory for Cell Biomechanics and Regenerative Medicine, Department of Biomedical Engineering, School of Life Science and Technology, Huazhong University of Science and Technology, Wuhan, Hubei 430074, China. <sup>2</sup>Department of Mechanical Science and Engineering, College of Engineering, University of Illinois at Urbana-Champaign, Urbana, Illinois 61801, USA. <sup>3</sup>Institute for Genomic Biology, University of Illinois at Urbana-Champaign, Urbana, Illinois 61801, USA. <sup>4</sup>Department of Bioengineering, University of Illinois at Urbana-Champaign, Urbana, Illinois 61801, USA. <sup>5</sup>Center for Biophysics and Computational Biology and Center for the Physics of Living Cells, Department of Physics, University of Illinois at Urbana-Champaign, Howard Hughes Medical Institute, Urbana, Illinois 61801, USA. <sup>6</sup>Department of Biochemistry and Molecular Biology, Tongji Medical College, Huazhong University of Science and Technology, Wuhan, Hubei 430030, China. <sup>7</sup>Department of Bioengineering and Institute of Engineering in Medicine, University of California-San Diego, La Jolla, California 92093, USA. <sup>8</sup>Department of Immunology, Institute of Basic Medical Sciences of Chinese Academy of Medical Sciences, Beijing 100005, China. \* These authors contributed equally to this work. † Present address: Center for Neuro-Medicine, Brain Science Institute, Korea Institute of Science and Technology, Hwarangno 14-gil 5, Seongbuk-gu, Seoul 136-791, South Korea. Correspondence and requests for materials should be addressed to N.W. (email: nwangrw@illinois.edu).

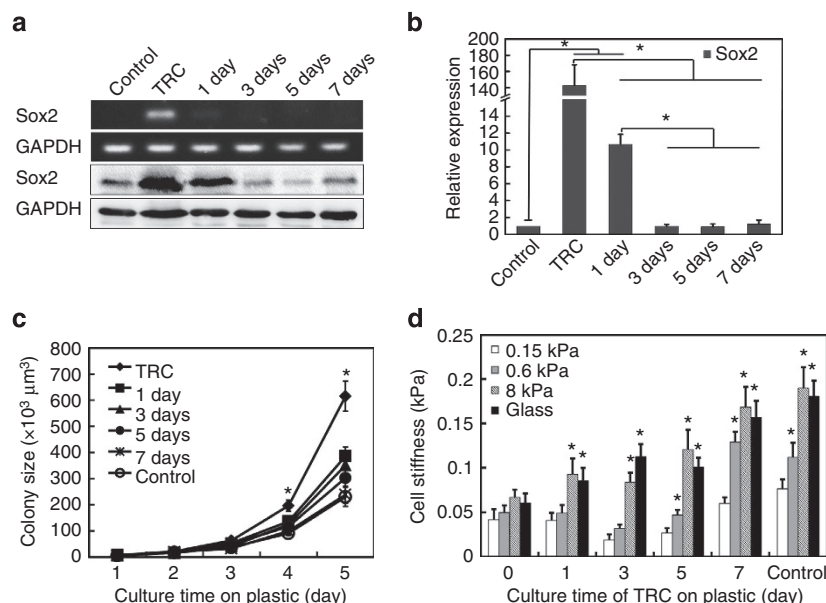
**T**umour-initiating cells (TICs) are a self-renewing, highly tumorigenic subpopulation of cancer cells. They play a critical role in the initiation and progression of cancer<sup>1</sup>. These tumorigenic cells exhibit high chemo-resistance to conventional chemotherapeutic drug treatment and therefore are speculated to be key players in cancer relapses after chemotherapy<sup>2</sup>. However, the concept of TICs has been controversial. Past reports show that a high percentage (>25%) of human melanoma cells can generate a tumour in a NOD-SCID interleukin-2 receptor- $\gamma$  chain null (IL2 $\gamma^{-/-}$ ) mouse<sup>3,4</sup>, suggesting that there is no hierarchy of clonal repopulation in melanoma. We recently developed a mechanical method of selecting TICs from cancer cell lines and primary cancer cells by culturing single cancer cells in soft fibrin gels<sup>5</sup>. Remarkably, in addition to being able to generate local primary tumours in wild-type syngeneic mice, when injected in tail veins, as few as ten of such cells can generate distant metastatic colonization and grow tumours in the lungs of wild-type non-syngeneic mice. Therefore, we functionally define these soft-fibrin-gel-selected melanoma cells as tumour-repopulating cells (TRCs) based on their high efficiency in repopulating tumours in wild-type syngeneic and non-syngeneic mice. Soon after our report, three other groups independently provide strong experimental evidence in mice that TRCs do exist in brain<sup>6</sup>, skin<sup>7</sup> and intestinal<sup>8</sup> tumours. *In vivo* imaging of unperturbed tumours further confirmed the existence of TRCs<sup>9,10</sup>. However, the underlying mechanisms of how TRCs maintain their self-renewing capability remain elusive. In the current study, we demonstrate that melanoma TRCs exhibit plasticity in mechanical stiffening, histone 3 lysine residue 9 (H3K9) methylation, Sox2 expression and self-renewal. Three-dimensional (3D) soft fibrin matrices promote

H3K9 demethylation and increase Sox2 expression and self-renewal, whereas stiff ones exert opposite effects.

## Results

**Self-renewal plasticity of TRCs.** It is known that soft substrates can sustain self-renewal of mouse embryonic stem cells<sup>11</sup> and substrate rigidity can regulate the fate of mesenchymal stem cells<sup>12</sup>, indicating that rigidity of extracellular matrix plays an important role in the maintenance and regulation of stem cell properties. As TRCs are selected from a population of melanoma cells that are usually cultured on rigid plastic, we asked what would happen if we plated these TRCs back to rigid substrates. To determine the effect of substrate rigidity on the gene expression, we cultured TRCs on rigid plastic for 1, 3, 5 and 7 days, and quantified their Sox2 expression. TRCs gradually lost Sox2 expression in both mRNA and protein levels along with the culture time on plastic (Fig. 1a,b and Supplementary Fig. 18c,d). Sox2 expression of TRCs dramatically decreased after 1 day and was as low as that of control cells after 3 days on plastic. Other stem cell genes *Bmi-1*, *C-kit*, *Nestin* and *Tert*, which are upregulated in TRCs<sup>5</sup>, were also downregulated after culture on plastic (Supplementary Fig. 1).

To examine the functional consequences of the loss of Sox2 and other stem cell genes, we re-plated those TRCs back into 90-Pa soft fibrin gels after culture on rigid substrates for 1, 3, 5 and 7 days, respectively. The growth rate of spheroids in fibrin matrices successively decreased with the culture time of TRCs on plastic (Fig. 1c), which is not a result of the increased apoptosis rate (Supplementary Fig. 2). Moreover, colony number was also decreased (Supplementary Fig. 3). After 7-day culture on plastic,



**Figure 1 | Inhibition of Sox2 expression and self-renewal of TRCs on 2D rigid substrates.** (a) Sox2 expression at both mRNA level (top panel) and protein level (bottom panel). Control: B16-F1 cells cultured on plastic. TRC: Control B16-F1 cells were cultured in 90-Pa fibrin gels for 5 days. 1 day, 3 days, 5 days or 7 days: TRCs were seeded on 2D rigid dishes for 1, 3, 5 or 7 days. Images are representatives of three independent RT-PCRs and two independent western blotting experiments. (b) Quantification of Sox2 expression by real time RT-PCR. Mean  $\pm$  s.e.m.;  $n=3$  independent experiments. \* $P<0.05$ . (c) 2D rigid substrates inhibit TRC growth. Significant differences between Control and TRC, or 1 or 3 days from Day 1 to 5 in the 90-Pa fibrin gels (all  $P<0.05$ , except TRC at day 2, where  $P=0.068$ , 1 day at day 2, where  $P=0.52$ , and 3 days at Day 2 and Day 3, where  $P>0.15$ ). No differences between Control and 5 or 7 days from Day 1–5 (all  $P>0.06$ , except 5 days at Day 1 and Day 3, where  $P<0.04$ ). Each data point was averaged from at least 30 colonies. \* $P<0.05$  between TRC and all other groups. (d) 2D rigid substrates promote stiffening of TRCs with substrate rigidity. B16-F1 cells cultured on plastic were used as a control. All were compared with '0.15 kPa' in each group. Mean  $\pm$  s.e.m.; \* $P<0.01$ . Each data point was averaged from >150 cells from two independent experiments. All the statistics were conducted using Student's *t*-test throughout the manuscript unless otherwise specified.

TRC colonies proliferated at a similar rate as control cells (harvested from rigid dishes). The lower colony size and number suggest that rigid substrates significantly reduce the self-renewal capability of TRCs, likely due to the loss of *Sox2* and other stem cell gene expressions.

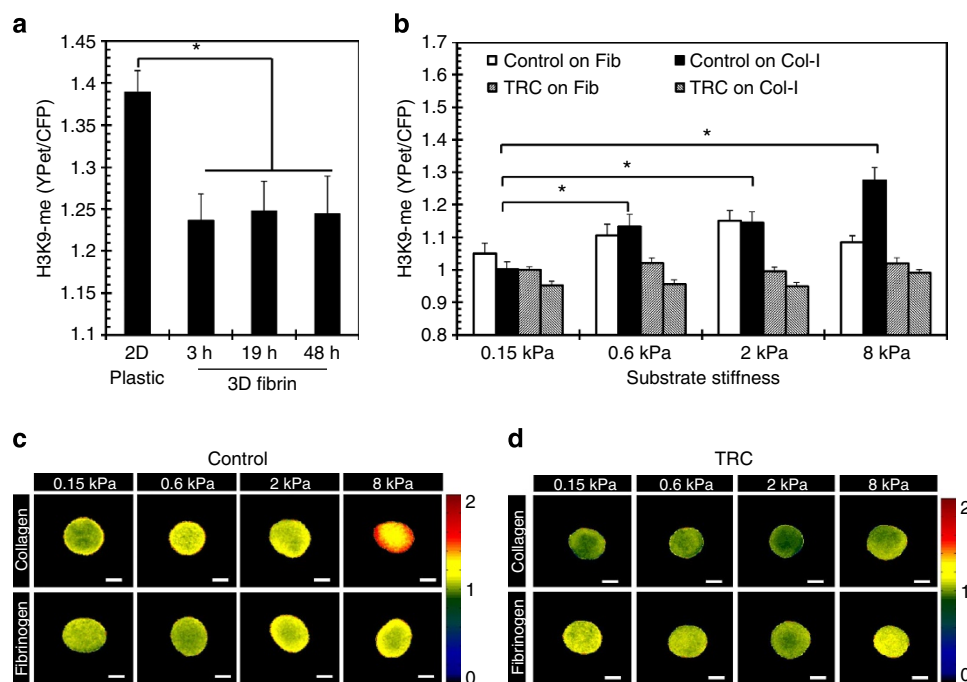
Besides the differences in gene expression patterns, TRCs also differ from control melanoma cells in their non-stiffening response to the increase in substrate rigidity<sup>5</sup>. To explore whether switching to rigid plastic may also change biophysical properties of TRCs, we first seeded them on two-dimensional (2D) rigid plastic for 1–7 days, and then quantified their stiffness on polyacrylamide (PA) gels of different rigidities. TRCs started to stiffen with substrate rigidity after 1-day culture on plastic (Fig. 1d). The stiffening response was more evident after 3 and 5 days. Interestingly, after 7-day culture on plastic, the stiffness and the stiffening response of TRCs were essentially the same as those of control melanoma cells (Fig. 1d).

### Regulation of H3K9 methylation by matrix and soluble factors.

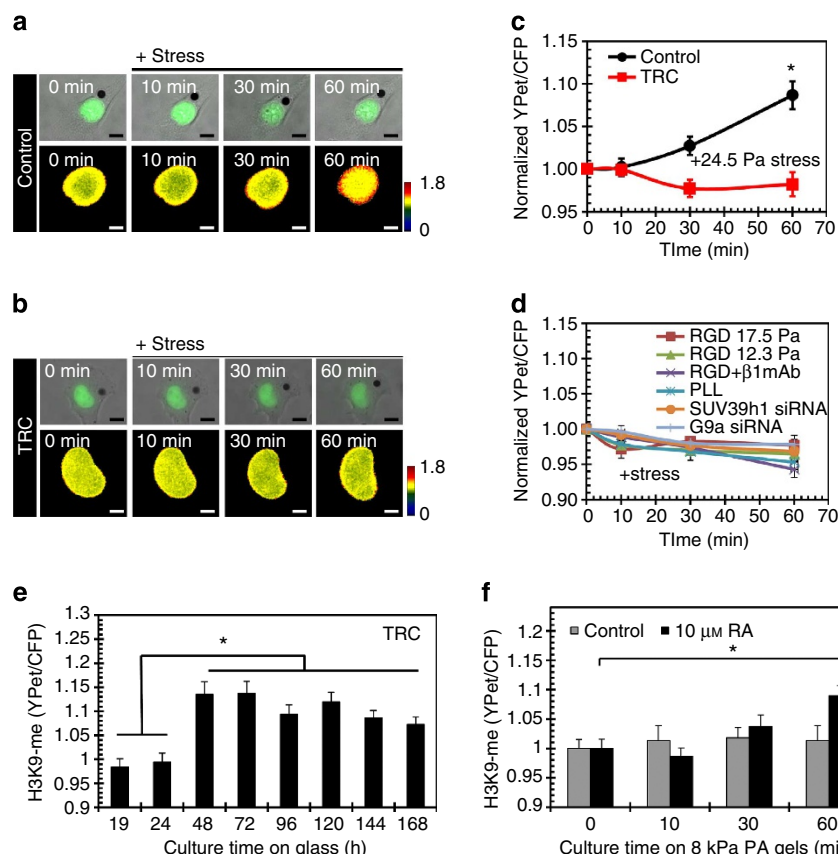
Our data have shown major differences in self-renewal gene expression and biophysical properties between TRCs and control melanoma cells. As these two types of cells are genetically identical, we hypothesized that epigenetics may play a role in it. A large body of work has shown aberrant changes in epigenetic features during tumour initiation and progression<sup>13,14</sup>, including histone modification. Among those aberrations, H3K9 is hypermethylated in several types of tumours<sup>15,16</sup>. We asked what role H3K9 methylation plays in TRCs. To explore H3K9 methylation of TRCs, we developed a CFP-YPet H3K9 fluorescence resonance energy transfer (FRET) biosensor (fused with a nuclear localization signal (NLS)) modified from a

previously reported CFP-YFP H3K9 biosensor<sup>17</sup> to quantify H3K9 di- and trimethylation (me2 and me3) in the nucleus of single cells (Supplementary Fig. 4a). This FRET probe uniquely measures the spatial distribution of H3K9 methylation level in the nucleus and revealed that H3K9 me2 and me3 mostly occurred at the nuclear periphery (Supplementary Fig. 4b). Culturing control cells in 90-Pa fibrin gels for merely 3 h resulted in H3K9 demethylation (Fig. 2a). TRCs did not elevate their H3K9 methylation (within 15 h of plating) with substrate stiffness on PA gels coated with either collagen-1 or fibrinogen (Fig. 2b,d); in contrast, control cells increased their H3K9 methylation with substrate stiffness on PA gels coated with collagen-1 but not with fibrinogen (Fig. 2b,c). Interestingly, high level of H3K9 methylation in control cells was associated with high cellular tractions on collagen-1-coated substrates and inhibition of the tractions decreased H3K9 methylation (Supplementary Fig. 5), suggesting that high endogenous forces transmitted via  $\alpha_1\alpha_2\beta_1$  (collagen-1 receptor) rather than via  $\alpha_v\beta_3$  (fibrinogen receptor) might be responsible for the stiff substrate-mediated H3K9 methylation, consistent with a report that activation of  $\beta_1$  integrin but not  $\beta_3$  integrin elevates cell traction forces<sup>18</sup>.

To further explore the role of physical forces in H3K9 methylation, we applied exogenous forces via RGD (Arg-Gly-Asp)-coated magnetic beads to single cells using magnetic twisting cytometry<sup>19</sup> (Fig. 3a,b). On fibrinogen-coated stiff substrates (Glass), the applied forces induced significant increases in H3K9 methylation at the nuclear periphery within 60 min only in control cells, but not in TRCs (Fig. 3a–c). The force-induced H3K9 methylation was dependent on force magnitude and specifically mediated via  $\beta_1$  integrin, as low forces, forces applied via beads coated with poly-L-lysine, or forces applied after blocking  $\beta_1$  integrin with monoclonal



**Figure 2 | Matrix rigidity mediated H3K9 methylation in control cells but not in TRCs.** (a) H3K9 methylation is lower in 90-Pa fibrin gels than on rigid plastic. Control B16-F1 cells were transfected with the H3K9 FRET methylation biosensor and then plated either on gelatin-coated rigid plastic (2D plastic) or in the 90-Pa soft 3D fibrin gel (3D Fibrin). H3K9 methylation was measured at the indicated times after cell plating. Mean  $\pm$  s.e.m.;  $n > 75$  cells for each value ( $*P < 0.001$ ). (b) H3K9 methylation in control B16-F1 cells (Control) but not in TRCs increases on stiff PA substrates coated with collagen I (Col-1) but not fibrinogen (Fib). PA, polyacrylamide. H3K9 methylation was measured 15 h after cell plating. Mean  $\pm$  s.e.m.;  $n > 40$  cells for each value ( $*P < 0.005$ ). Representative FRET images of a cell nucleus for control cells (c) and TRCs (d) on PA gels coated with collagen I (top panel) or fibrinogen (bottom panel). Scale bars, 5  $\mu$ m.



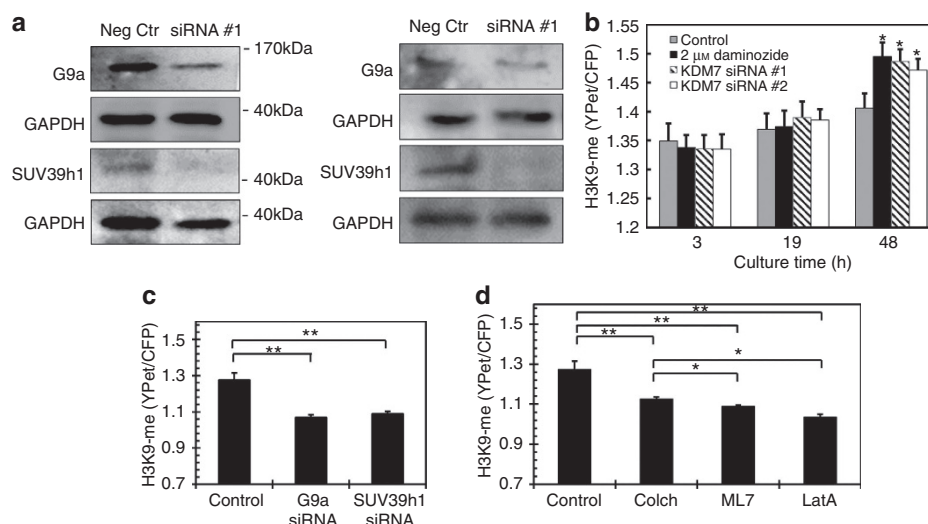
**Figure 3 | Exogenous forces increase H3K9 methylation level via  $\beta 1$  integrin.** Exogenous forces increase H3K9 methylation in control B16-F1 cells (**a**) but not in TRCs (**b**). Top: bright-field and fluorescence overlay image of the cells; the magnetic bead is shown as a black dot. Bottom: representative FRET images showing the spatial distribution of H3K9 methylation in the nucleus during loading. Cells were plated on 8 kPa PA gels coated with fibrinogen and loaded at 24.5 Pa, 0.3 Hz with RGD-coated ferromagnetic beads. H3K9 methylation clearly increases in **a** at the nuclear periphery 30–60 min after loading. Scale bars, 10  $\mu$ m (top), 5  $\mu$ m (bottom). (**c**) Quantification of H3K9 methylation in **a** and **b** during loading. (**d**) No force-induced elevation of H3K9 methylation was observed when the externally applied stress magnitude was equal or  $<17.5$  Pa, when  $\beta 1$  integrin was blocked by a monoclonal antibody, when the force was applied via a poly-L-lysine coated bead, or when siRNA was transfected to knockdown G9a and SUV39h1, which are methyltransferases responsible for dimethylation and trimethylation of H3K9, respectively. Knockdown of G9a and SUV39h1 was confirmed by western blottings that these proteins reduced by  $\sim 50\%$  when compared with control. (**e**) H3K9 methylation in TRCs increases after 2-day culture on glass dishes. TRCs were cultured on collagen I-coated glass-bottomed dishes and H3K9 methylation was measured at indicated times. Mean  $\pm$  s.e.m.,  $n > 40$  cells for each time point,  $*P < 0.05$ . (**f**) Short time treatment with retinoic acid (RA) increases H3K9 methylation in TRCs. TRCs were cultured on collagen I-coated 8-kPa PA gels for 15 h. Cells were then treated with or without (Control) 10  $\mu$ M RA. Note that all the values were normalized to the value at 0 min (15 h after plating). Mean  $\pm$  s.e.m.,  $n > 40$  cells for each value,  $*P < 0.05$ .

antibody, was not able to induce H3K9 methylation (Fig. 3d). In addition, silencing H3K9 methyltransferase G9a or SUV39h1 (knockdown efficiency was  $\sim 50\%$ ; Fig. 4a) not only reduced the baseline level of H3K9 methylation in control melanoma cells (Fig. 4c), but also completely blocked the force-induced H3K9 methylation (Fig. 3d), consistent with a report that these two enzymes are critical to H3K9 methylation<sup>20</sup>. In contrast, silencing or inhibiting H3K9 demethylase KDM7 with small interfering RNA (siRNA) or with Daminozide in cells in 3D soft fibrin gels increased H3K9 methylation levels (Fig. 4b). These data suggest that H3K9 methylation levels in these melanoma cells are controlled by both methyltransferases and demethylases. H3K9 methylation also depended on intactness of the cytoskeleton and cellular tension, as disrupting actin microfilaments with Latrunculin A or microtubules with colchicine and inhibiting myosin light chain kinase with ML7 significantly lowered H3K9 methylation levels (Fig. 4d). Although TRCs did not increase H3K9 methylation with substrate stiffness within 15 h (Fig. 2b,d), they started to increase H3K9 methylation on collagen-1-coated plastic after 48 h (Fig. 3e). In contrast, TRCs treated with retinoic

acid (a nonspecific soluble differentiation factor) for 60 min (similar in duration for external force-induced H3K9me; see Fig. 3c) increased H3K9 methylation levels (Fig. 3f), suggesting H3K9 methylation may be associated with the differentiation of TRCs.

**H3K9 demethylation-induced Sox2 expression is critical to TRC self-renewal.** As H3K9 methylation has been associated with inactivation of certain genes<sup>13–15</sup>, we postulated that H3K9 demethylation might be related to Sox2 expression. Our western blotting analysis showed that control cells expressed higher levels of H3K9 me2 and H3K9 me3 compared with TRCs (Supplementary Fig. 6a). Importantly, silencing H3K9 methyltransferase G9a or SUV39h1 in control cells significantly increased their Sox2 expression (Fig. 5a and Supplementary Fig. 7a), suggesting that H3K9 demethylation can increase Sox2 expression. To further investigate how H3K9 demethylation promotes Sox2 expression, we performed chromatin immunoprecipitation (ChIP) assay to quantify H3K9 me2 and me3 levels





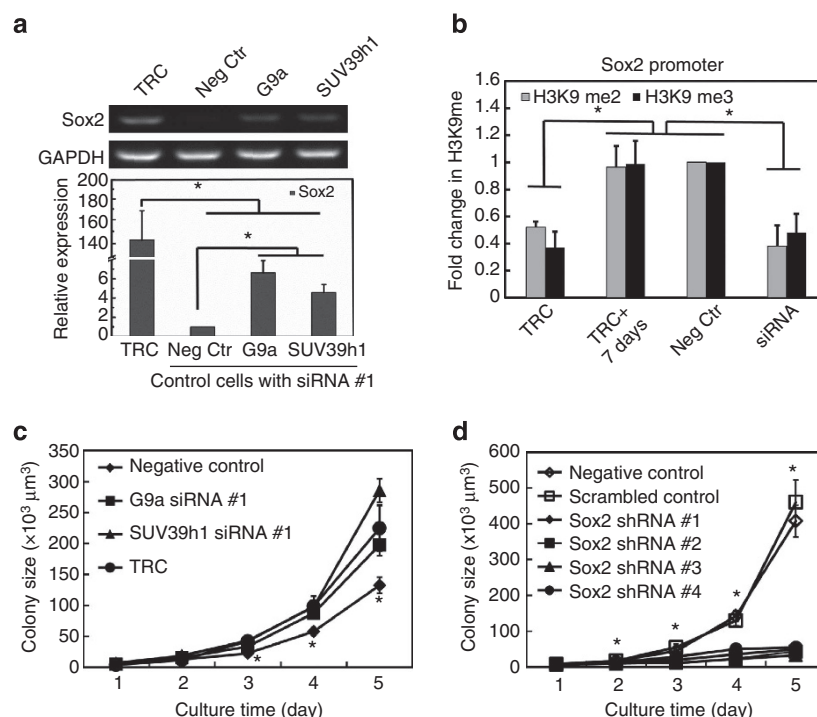
**Figure 4 | H3K9 methylation levels depend on methyltransferases and demethylases and the tensed cytoskeleton.** (a) Representative images of western blotting of G9a or SUV39h1 expression after siRNA knockdown. Control melanoma cells were transfected with two sets of G9a/SUV39h1 siRNAs (1 and 2), respectively, and western blotting assays were performed to confirm the knockdown efficiencies to be 48% (1 siRNA) and 45% (2 siRNA) for G9a, and 58% (1 siRNA) and 43% (2 siRNA), respectively. Three independent experiments showed similar results. (b) Silencing or inhibiting demethylase KDM7 increases H3K9 methylation of control melanoma cells in soft fibrin gels. Control cells were transfected with H3K9 FRET biosensors and KDM7 siRNAs (1 and 2) or negative control siRNA, or treated with KDM7 inhibitor daminozide. The cells were seeded into 3D soft fibrin gels and H3K9 methylation levels were measured after 3, 19 or 48 h. Mean  $\pm$  s.e.m.;  $n > 40$  cells for each data point;  $*P < 0.05$ . (c) Knockdown of G9a or SUV39h1 decreases the H3K9 methylation in control B16-F1 cells. (d) Inhibiting microtubule, myosin light chain kinase (MLCK) or actin polymerization decreases H3K9 methylation. Control B16-F1 cells were incubated with Colchicine (Colch, 10  $\mu$ M for 30 min) to inhibit microtubule polymerization, ML7 (25  $\mu$ M for 30 min) to inhibit myosin light chain kinase, or Latrunculin A (LatA, 1  $\mu$ M for 30 min) to disrupt actin filaments. Mean  $\pm$  s.e.m.;  $n > 40$  cells for each data point;  $*P < 0.05$ ;  $**P < 0.001$ .

in the promoter region of *Sox2*. H3K9 me2 and H3K9 me3 levels were  $\sim 50\%$  higher in the promoter region of *Sox2* in control cells than in TRCs (Fig. 5b and Supplementary Fig. 6b). Silencing G9a or SUV39h1 in control cells greatly decreased their H3K9-me2 or H3K9-me3 levels in the promoter region of *Sox2* (Fig. 5b). Plating TRCs on plastic for 7 days elevated H3K9 me2 and me3 to the same levels as those in control cells (Fig. 5b and Supplementary Fig. 6b). These results suggest that G9a and SUV39h1 methylated H3K9 at the *Sox2* promoter site blocks *Sox2* expression. In contrast, TRCs did not express G9a and SUV39h1 proteins and silencing *Sox2* in TRCs did not increase the expression of G9a or SUV39h1 (Supplementary Fig. 8), suggesting that G9a and SUV39h1 may be independent of *Sox2* expression.

As knocking down G9a or SUV39h1 can induce *Sox2* expression in control cells, we further explored whether H3K9 demethylation can promote their self-renewal capability. Silencing G9a or SUV39h1 in control cells significantly increased the colony growth in soft fibrin gels to a level similar to that of TRCs (Fig. 5c and Supplementary Fig. 7b), but the colony number was still twofold less than that of TRCs (Supplementary Fig. 7c,d). However, simultaneously silencing G9a (or SUV39h1) and *Sox2* completely abrogated the colony growth but not the colony number (Supplementary Fig. 9), suggesting that H3K9 demethylation promotes self-renewal capability of melanoma cells in a *Sox2*-dependent manner. H3K9 methylation is regulated by both methyltransferases and demethylases. Silencing KDM7, an important demethylase for H3K9 (ref. 21), significantly elevated H3K9 methylation (Fig. 4b) and suppressed colony growth and number (Fig. 6a,b), probably through inhibition of *Sox2* expression (Fig. 6c). Treatment with two separate inhibitors of KDM4 (ref. 22) (another demethylase for H3K9) led to dose-dependent inhibition of colony size and colony number (Supplementary Fig. 10). Taken together, these data suggest that H3K9 demethylation by deactivating methyltransferases or

activating demethylases increases *Sox2* expression and thus promotes self-renewal capability.

We further investigated the functional role of *Sox2* in maintaining self-renewal of TRCs *in vitro* and *in vivo*. Silencing *Sox2* greatly decreased the expressions of *Sox2*, *Bmi-1* and *C-kit* in TRCs (Supplementary Fig. 11). Colony growth was suppressed by  $> 90\%$  in *Sox2*-knocked down TRCs in comparison with that of control TRCs (Fig. 5d), suggesting that *Sox2* is essential in maintaining self-renewal of TRCs *in vitro*. *Sox2* silencing had no effects on control melanoma cells (these cells express little *Sox2*) that were grown on plastic and were then embedded in 3D stiff fibrin gels, suggesting that there were little off-target effects of *Sox2* silencing (Supplementary Fig. 12). Overexpressing *Sox2* in control melanoma cells increased the colony number but not colony growth (Supplementary Fig. 13), indicating that this intervention increases the number of self-renewing cells but does not alter cell cycle duration of a self-renewing cell. Importantly, subcutaneously injecting 100 of TRCs treated with *Sox2* short hairpin RNA (shRNA) into wild-type syngeneic C57BL/6 mice did not generate any tumours by day 19. In contrast, palpable tumours were observed for untreated TRCs by day 13 and treated with scrambled control shRNA by day 15. By day 28, 2 out of 16 mice for TRCs treated with *Sox2* shRNA had small tumours (Supplementary Table 1), whereas for untreated TRCs 9 out of 16 mice formed large tumours; for TRCs treated with scrambled shRNA, 3 out of 8 formed tumours. The formation of tumours in TRCs treated with *Sox2* shRNA appearing at late stages probably reflects the fact that the knockdown of *Sox2* by shRNA was transient such that *Sox2* expression came back in these cells after shRNA effect was depleted. As expected from previous findings<sup>5</sup>, injecting 100 of control melanoma cells (harvested from rigid plastic) did not form any tumours (0/8; Supplementary Table 1). These data demonstrate that *Sox2* expression is necessary to sustain self-renewal capability of TRCs both *in vitro* and *in vivo*.



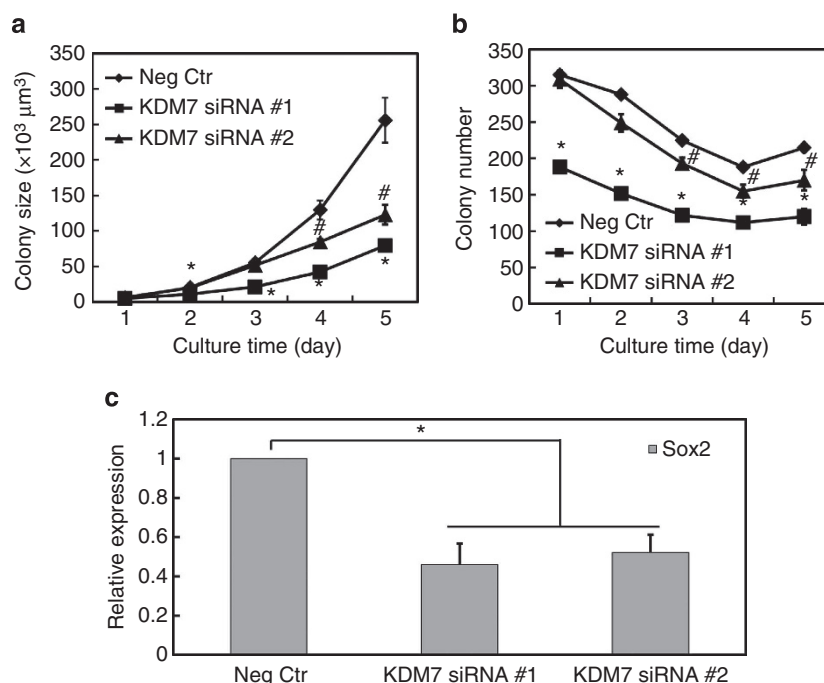
**Figure 5 | H3K9 demethylation-mediated Sox2 expression is essential for *in vitro* tumour growth.** (a) Silencing G9a or SUV39h1 promotes Sox2 expression in control melanoma cells, quantified by real-time RT-PCR. Neg Ctr: negative control. \* $P < 0.05$ . (b) H3K9 di- (me2) and trimethylation (me3) levels in the promoter region of Sox2 quantified by the ChIP assay. TRC + 7 days: TRCs were cultured on plastic for 7 days; Neg Ctr, siRNA: control melanoma cells were treated with negative control siRNA, G9a (for H3K9 me2) or SUV39h1 (for H3K9 me3) siRNA #1, respectively. \* $P < 0.05$ . (c) H3K9 demethylation in control melanoma cells promotes their self-renewal capability. Control cells were transfected with G9a or SUV39h1 siRNA and then re-plated in soft fibrin gels. (d) Silencing Sox2 greatly inhibits growth of TRC spheroids. TRCs were transfected with Sox2 shRNAs or scrambled shRNA. These cells were then re-plated into 90-Pa fibrin gels. Each data point was averaged from at least 40 colonies. In c, significant differences between Negative Control and G9a siRNA 1 or SUV39h1 siRNA 1 or TRC from day 2 to day 5 (\* $P < 0.05$ , except G9a siRNA 1 at day 2, where  $P = 0.09$ ). In d, significant differences between Negative control or Scrambled control and Sox2 shRNA 1 to 4 from day 2 to day 5. (\* $P < 0.05$ ). Mean  $\pm$  s.e.m.;  $n = 3$  independent experiments for all subfigures.

Because of the importance of Sox2 and H3K9 demethylation in TRCs, we asked what roles Sox2 expression and H3K9 methylation might play in the selection and propagation of TRCs by soft fibrin gels. When control melanoma cells were transfected with Sox2 shRNA and cultured in 90-Pa fibrin gels, both growth and number of spheroids were inhibited (Supplementary Fig. 14a,b). Moreover, treatment with Daminozide, a highly selective inhibitor of KDM7, significantly inhibited TRC spheroid growth number (Supplementary Fig. 14c,d). Therefore, these data suggest that H3K9 demethylation-dependent regulation of Sox2 expression is important in TRC selection and propagation.

**Mechanism of cell stiffening with substrate stiffness.** To elucidate the mechanisms of the stiffening response plasticity, we examined the potential role of Cdc42 in cell stiffening because of its known role in inducing filopodia formation and cell spreading in B16-F1 melanoma cells<sup>23</sup>. TRCs expressed ~50% less Cdc42 than control cells; plating TRCs on plastic for 5 and 7 days increased their Cdc42 expression by ~30% and ~100%, respectively (Fig. 7a). Overexpression of Cdc42 in TRCs rescued the cell stiffening response with substrate rigidity (Fig. 7b), whereas silencing Cdc42 in control melanoma cells or in TRCs on plastic for 5 days completely abolished the stiffening response (Fig. 7c). Silencing Cdc42 in melanoma cells on rigid plastic downregulated H3K9 me2 but not H3K9 me3 (Fig. 7d and Supplementary Fig. 15a,b) and had no effects on Sox2 expression (Supplementary Fig. 15c), suggesting that Cdc42, cell stiffening

and H3K9 methylation might be linked via tension-dependent cytoskeletal mechanotransduction, contributing to the biological plasticity of TRCs.

**Inhibition of TRC self-renewal by 3D stiff fibrin matrix.** When the cells were cultured from 2D rigid dish to 3D soft matrix, both matrix dimensionality and stiffness were altered. To further determine whether it is the 3D matrix stiffness or the effect of switching from 2D to 3D matrix that impacts on the self-renewal capability of TRCs, we cultured control B16-F1 cells in 3D soft (90-Pa) or 3D stiff (1,050-Pa) fibrin gels for 5 days. These cells were then seeded into either soft or stiff fibrin gels. After re-seeding, the cells grew better in soft fibrin gels no matter whether they were derived from soft or stiff fibrin gels; in contrast, once re-seeded in stiff fibrin gels, colony growth and number were inhibited (Fig. 8a,b), suggesting that switching from a 3D soft matrix to a 3D stiff matrix inhibits self-renewal and promotes quiescence of the TRCs. When culturing control melanoma cells in soft and stiff fibrin gels, a 10-fold increase in substrate stiffness leads to a ~1.25-fold increase in H3K9 me2 and a 50% increase in H3K9 me3 on the protein level, and a ~90% decrease in Sox2 expression (Fig. 9). Importantly, switching from a stiff matrix (1,050-Pa) to a soft matrix (90-Pa) significantly decreased H3K9 methylation and elevated Sox2 expression by ~5-fold (Fig. 8c,d). Taken together, these data suggest that 3D stiff fibrin matrix can inhibit TRC self-renewal via H3K9 methylation-mediated Sox2 suppression.



**Figure 6 | Silencing H3K9 demethylase KDM7 greatly inhibits colony size and number through suppression of Sox2 expression.** Knocking down KDM7 using two different siRNAs significantly inhibits colony size (**a**) and number (**b**). Control melanoma cells were transfected with KDM7 siRNA 1 or 2, and then cultured in 90-Pa fibrin gels. Colony number was counted and colony size was monitored until day 5 ( $n=2$ ). There is a significant difference between Neg Ctr and KDM7 siRNA 1 in **a** from day 2 through day 5 (all  $P<0.00001$ ) and in **b** from day 1 through day 5 (all  $P<0.01$ ), and between Neg Ctr and KDM7 siRNA 2 in (**a**) at day 4 and day 5 (both  $P<0.005$ ) and in **b** from day 3 through day 5 (all  $P<0.05$ ). '\*' and '#' indicates significant differences between Neg Ctr and KDM7 siRNA 1 or 2, respectively. (**c**) Silencing KDM7 significantly suppresses Sox2 expression. Control melanoma cells were transfected with KDM7 siRNA 1 or 2, or negative control siRNA and then cultured in 90-Pa fibrin gels. After 5 days, Sox2 expression was quantified by real time RT-PCR. \* $P<0.05$ ;  $n=3$  independent experiments.

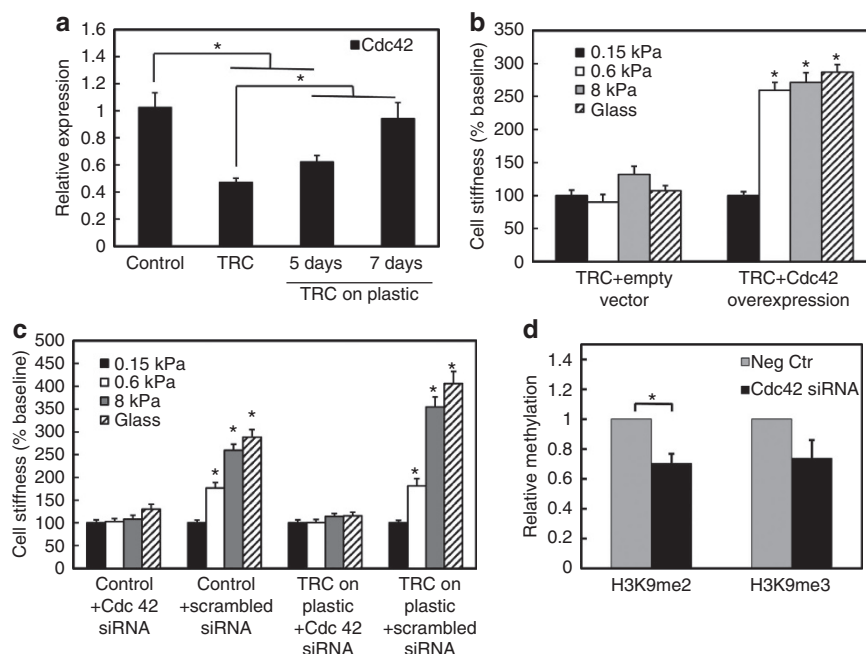
## Discussion

Increasing evidence has demonstrated that TRCs truly exist as a highly tumorigenic subpopulation that is critical to tumour initiation and progression. TRCs also exhibit high resistance to conventional chemotherapeutic drug treatment, which has been speculated to drive cancer relapse after chemotherapy. Therefore, TRCs may be one of the pivotal targets to achieve complete tumour eradication<sup>24</sup>. In the present study, we investigated how TRCs maintain their self-renewal capability. As TRCs appear to be less differentiated, we set out to determine their specific lineage. Our data show that TRCs exhibited *Sox2*<sup>high</sup>*Mitf*<sup>low</sup> phenotype and control melanoma cells exhibited *Sox2*<sup>-</sup>*Mitf*<sup>+</sup> phenotype (Supplementary Fig. 16a,b), consistent with published results<sup>25–27</sup> and suggesting that TRCs resemble melanoblasts that are *Sox2*<sup>low</sup>*Mitf*<sup>low</sup> and control melanoma cells resemble melanocytes that are *Sox2*<sup>-</sup>*Mitf*<sup>+</sup> in these two gene expression patterns<sup>27</sup>. However, expression of *Mitf* upstream regulators, *Pax3* (ref. 28) and *Sox10* (ref. 27), and melanocyte-specific genes, tyrosinase-related protein 1 (*Tyrp1*)<sup>28</sup>, dopachrome tautomerase (*DCT*)<sup>28</sup> and glycoprotein non-metastatic melanoma protein B (*Gpnmb*)<sup>29</sup>, was similar in TRCs as in control cells (Supplementary Fig. 16c,d). These findings suggest that melanoma TRCs may be progenitor cells of control B16-F1 cells but more differentiated than melanoblasts. Moreover, knocking down *Sox2* in TRCs increased *Mitf* expression (Supplementary Fig. 17a), while silencing *Mitf* (knockdown efficiency ~80%; Supplementary Fig. 17b) in control melanoma cells did not affect *Sox2* expression (Supplementary Fig. 17c), suggesting that self-renewal gene *Sox2* is independent of *Mitf*. Importantly, when TRCs are plated on the rigid dish, expression of melanocyte-related gene *Mitf* gradually increased (Supplementary Fig. 18), demonstrating *Mitf* plasticity

of TRCs when substrate rigidity was elevated, opposite in direction as the plasticity of *Sox2* expression (see Fig. 1a,b).

*Sox2* is known for its critical role in the maintenance of pluripotency<sup>25</sup> and fate determination<sup>30</sup> of stem cells. *Sox2* also sustains self-renewal capability of TICs<sup>26</sup>. Inhibition or silencing of *Sox2* leads to loss of tumorigenicity<sup>26,31</sup>. *Sox2* dysfunction in cancer<sup>32,33</sup> is known, but it is not clear what triggers *Sox2* expression in cancer. Our current study reveals a critical role of the mechanics of 3D matrix in *Sox2* expression in a subpopulation of tumour cells. Importantly, silencing *Sox2* greatly inhibits spheroid growth *in vitro* and tumour generation *in vivo*. TICs are heterogeneous in terms of their transient, long-term, or delayed-contributing effects on tumour growth<sup>34</sup>. Although we cannot exclude the possibility that TRCs isolated from soft fibrin matrices are also heterogeneous, our previous finding that as few as ten of these TRCs are sufficient to generate local and distant tumours<sup>5</sup> suggests that these TRCs are distinct from those TICs.

Cancer cells exhibit aberrant epigenetic features, including global changes in DNA methylation and altered histone modification<sup>35</sup>. On the other hand, stem cells have an open chromatin structure and abnormal epigenetic regulation may be prone to malignant transformation<sup>36</sup>. Here we show that H3K9 demethylation by silencing G9a or SUV39h1 can greatly promote the self-renewal capability of control melanoma cells in a *Sox2*-dependent manner, whereas silencing H3K9 demethylases inhibits self-renewal. It should be noted that in contrast to the increase in TRC colony number and growth observed following culture in soft substrates, silencing H3K9 methyltransferases has significant functional effects on colony growth but not colony number and the levels of *Sox2* remain 30- to 40-fold lower than those found in TRCs. The exact mechanism is not clear at this



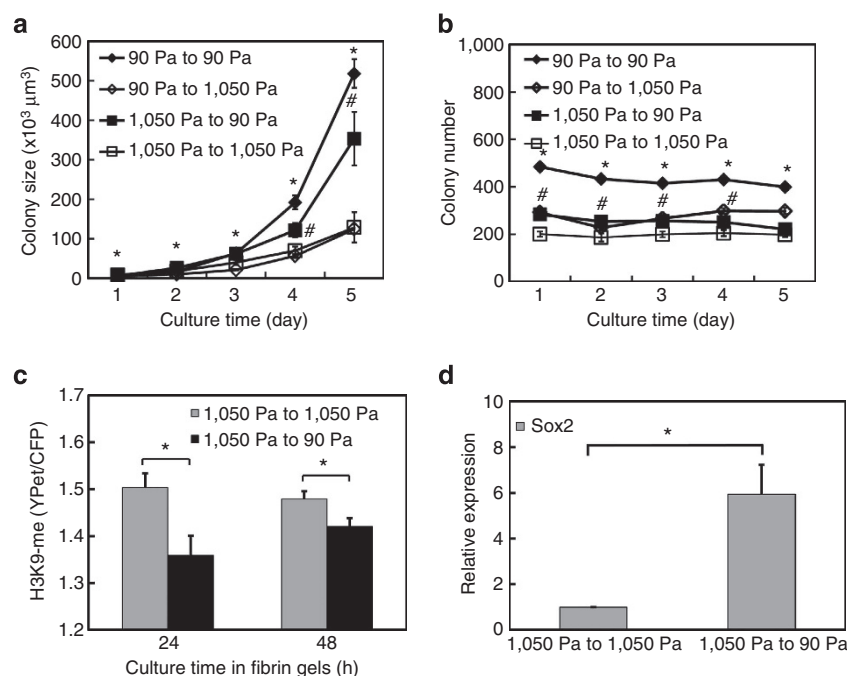
**Figure 7 | *Cdc42* expression regulates stiffening response and H3K9 me2 in melanoma cells.** (a) TRCs express lower level of *Cdc42* compared with control melanoma cells and TRCs after culture on rigid plastic for 5 or 7 days.  $n = 6$ ;  $P < 0.05$ ; two independent experiments. (b) TRCs regain cell stiffening responses after overexpression of *Cdc42*. Cells were plated on Collagen-I-coated PA gels with different stiffnesses or on glass-bottomed dishes. Experiments were carried out 6 h after plating. Relative changes in cell stiffness were represented with respect to the cell stiffness on 0.15 kPa PA gel as the baseline. TRC + empty vector: TRCs were transfected with pEGFP-N1 vector; TRC + *Cdc42* overexpression: TRCs were transfected with *Cdc42*-EGFP plasmid.  $n > 150$  cells;  $P < 0.001$ . (c) *Cdc42* knockdown inhibits cell stiffening in control cells and TRCs plated on plastic for 5 days. Relative changes in cell stiffness were represented with respect to the cell stiffness on 0.15 kPa PA gel as the baseline.  $n > 100$  cells;  $P < 0.001$ . Control: B16-F1 cultured on rigid plastic; TRC on plastic: TRCs were cultured on plastic for 5 days; Control + *Cdc42* siRNA: control melanoma cells were transfected with *Cdc42* siRNA; Control + Scrambled siRNA: control cells were transfected with scrambled siRNA; TRC on plastic + *Cdc42* siRNA: TRCs were cultured on plastic for 5 days and then transfected with *Cdc42* siRNA; TRC on plastic + Scrambled siRNA: TRCs were cultured on plastic for 5 days and then transfected with scrambled siRNA. Mean  $\pm$  s.e.m.; '\*' indicates significant difference compared with '0.15 kPa'. (d) Quantification analysis of western blotting of H3K9me2 and me3 after *Cdc42* knockdown in control melanoma cells. Mean  $\pm$  s.e.m.;  $n = 3$  independent experiments;  $P < 0.05$ .

time, but it may be partly due to the fact that only 50% of G9a/SUV39h1 is knocked down, whereas there are no detectable levels of G9a/SUV39h1 in TRCs (Supplementary Fig. 8) and/or other self-renewal promoting effects of silencing G9a/SUV39h1 are at work. In addition, it is not clear why silencing *Sox2*, overexpressing *Sox2*, or inhibiting KDM4, has different effects from silencing methyltransferases on colony number and size. These issues need to be investigated in the future by dissecting out specific signalling pathways. Nevertheless, our data suggest that H3K9 demethylation and methylation, modulated by force-induced chromatin remodelling mediated by the cytoskeleton, can regulate *Sox2* expression and play important roles in self-renewal and differentiation of TRCs. Our results show that stiff matrix induced H3K9 methylation occurs at the nuclear periphery just under the nuclear envelope. The applied stress (via the magnetic bead) on the cell surface via integrins also induces H3K9 methylation at the periphery of the nucleus, suggesting that stiff matrix-induced H3K9 methylation may be (at least) in part due to the elevation of endogenous stresses in the cell. These findings are in line with a recent report that force-induced stiffening of isolated nuclei depends on an intact LINC (linker of nucleoskeleton and cytoskeleton) pathway<sup>37</sup>. *Cdc42* silencing decreases H3K9me2 by  $\sim 20\%$ , similar to that after disruption of cytoskeletal filaments with drugs (Fig. 4d), suggesting that the effect of silencing *Cdc42* might be important in altering cell functions. However, silencing *Cdc42* has no effect on H3K9me3 or *Sox2*, suggesting that some molecule(s) upstream of *Cdc42* may be important in controlling H3K9me3 and *Sox2*.

Furthermore, our current findings suggest that increasing matrix rigidity affect *Cdc42*-mediated cell stiffening, H3K9 methylation and *Sox2* suppression, but whether these are independent or linked pathways in controlling TRC function remains unclear and requires additional experiments in the future. Published reports suggest that G9a/SUV39h1-mediated H3K9 methylation contributes to cancer initiation and/or progression<sup>13,14,16,38,39</sup>, different from our findings that G9a or SUV39h1 knockdown leads to *Sox2* expression. This discrepancy may lie in the fact that in those previous studies differentiated cancer cells are assayed, whereas in our present study TRCs are analysed. Although there are reports on *Sox2* expression in several types of undifferentiated tumour cells<sup>26,31</sup>, there is, to the best of our knowledge, no report of H3K9 methylation or *Sox2* expression in TRCs when mechanical microenvironment is perturbed. Therefore, our findings on low force-mediated H3K9 demethylation and thus *Sox2* gene expression are novel. Furthermore, we find that H3K9 methylation can directly inhibit *Sox2* expression. Nevertheless, the potential roles of other histone modifications, such as H3K4 and/or H3K27 methylation, acetylation and phosphorylation, and DNA methylation, remain to be elucidated. In addition, the mechanisms of *in vivo* *Sox2* expression/suppression with the link to H3K9 demethylation/methylation and matrix compliance/cell stiffening need to be investigated in the future.

We show that a uniform stiff fibrin gel functions as a physical barrier to prevent the formation and growth of TRC spheroids through elevated H3K9 methylation-induced *Sox2* suppression, supporting a recent postulate on breast cancer progression<sup>40</sup>.





**Figure 8 | Stiff matrix inhibits self-renewal of TRCs.** Stiff fibrin gels inhibit colony growth (a) and number (b). ‘90-Pa to 90-Pa’ and ‘90-Pa to 1,050-Pa’: control B16-F1 cells were cultured in 90-Pa fibrin gels for 5 days and these cells were then re-plated into 90-Pa and 1,050-Pa fibrin gels, respectively. ‘1,050-Pa to 90-Pa’ and ‘1,050-Pa to 1,050-Pa’: control B16-F1 cells were cultured in 1,050-Pa fibrin gels for 5 days and these cells were then re-plated into 90-Pa or 1,050-Pa fibrin gels. Spheroid size and number were measured from Day 1 through Day 5. In a and b, significant differences between ‘90-Pa to 90-Pa’ and ‘90-Pa to 1,050-Pa’ from Day 1 through Day 5 (all  $P < 0.01$ ); significant differences between ‘1,050-Pa to 90-Pa’ and ‘1,050-Pa to 1,050-Pa’ at day 4 and day 5 in a (both  $P < 0.02$ ) and from day 1 to day 4 in b (all  $P < 0.05$ ). Mean  $\pm$  s.e.m.;  $n = 2$  independent experiments. Each data point was averaged from  $> 30$  colonies. ‘\*’ and ‘#’ indicates significant differences between ‘90-Pa to 90-Pa’ and ‘90-Pa to 1,050-Pa’ and between ‘1,050-Pa to 90-Pa’ and ‘1,050-Pa to 1,050-Pa’, respectively. (c) Switching to 3D soft matrix decreases H3K9 methylation. Mean  $\pm$  s.e.m.;  $n > 50$  cells from two independent experiments;  $*P < 0.05$ . (d) Switching to 3D soft matrix increases Sox2 expression. Mean  $\pm$  s.e.m.;  $n = 3$  independent experiments;  $*P < 0.05$ . In c and d, control cells were cultured in 1,050-Pa fibrin gels for 5 days and then re-plated into 90-Pa or 1,050-Pa gels. After 24 or 48 h in c and 5 days in d, their H3K9 methylation was measured by FRET and Sox2 expression was quantified by real time RT-PCR.

Melanoma cells in the same soft fibrin gels can form round colonies only at z-heights far away from the rigid plastic bottom (Supplementary Fig. 19); as these cells experience the same porosity but different elasticity, these data demonstrate that it is matrix stiffness but not porosity that dictates round colony generation and self-renewal. Furthermore, addition of soluble fibrinogen ligands to the 90-Pa fibrin matrix (total ligand amount is equivalent to that of 1,050-Pa fibrin gels) did not alter colony growth (Supplementary Fig. 20), suggesting that the difference in colony growth between 90-Pa and 1,050-Pa fibrin gels may not be due to ligand density availability. It is noteworthy that the H3K9 methylation results and the corresponding traction data (and their inhibition by myosin light chain kinase inhibitor ML7) suggest that different integrin subsets (that is, collagen-1-mediated  $\beta 1$  integrin binding versus fibrinogen-mediated  $\beta 3$  integrin binding) can trigger different responses in H3K9 methylation even for the same substrate stiffness.

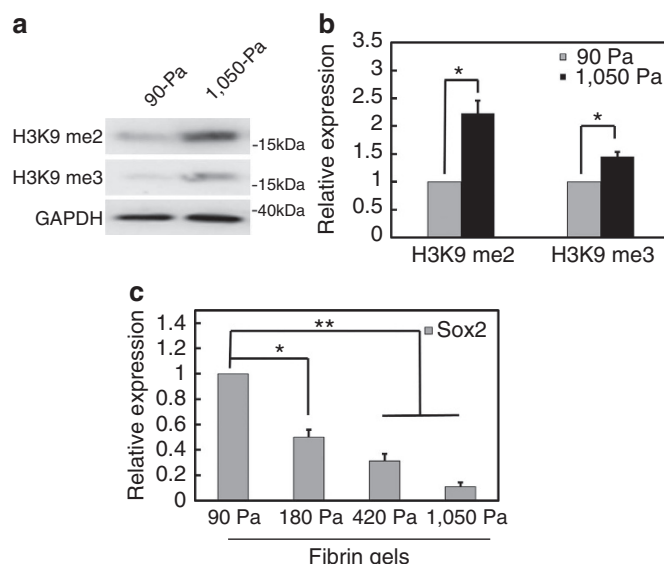
Our data suggest that TRCs may have different cell fates in matrices of various mechanical and biochemical microenvironments (Fig. 10). In a soft matrix, TRCs can proliferate well and grow into a local tumour. In the presence of soluble differentiation factors, TRCs cannot proliferate even in soft matrix. In contrast, a stiff matrix restricts TRC growth and makes them quiescent. However, on matrix degradation and remodelling, the stiff matrix may become locally softened, reactivating TRCs to proliferate, while some areas of local stiffening may promote TRCs to differentiate. Locally switching from 3D stiff matrices to 3D soft matrices and vice versa may be able to regulate TRC self-

renewal, as suggested by our current results from *in vitro* culture. This scenario may partially explain why tumour relapses several years after surgery and therapies. These data indicate that stiff matrix may be used as an effective strategy to suppress self-renewal of TRCs.

Published reports have shown that tension-dependent matrix stiffening is critical in breast cancer progression<sup>41–43</sup>. At a first look, it appears that there is a discrepancy in our findings and the results of these reports. A close examination reveals that our findings and theirs may be referring to different cancer cells at different stages during cancer progression. It is known that tissue stiffening alone cannot drive mammary epithelial transformation without ErbB2 activation<sup>43</sup>. It is possible that integrin-mediated, tension-dependent matrix stiffening facilitates differentiation and invasion of those TRCs only in the presence of matrix degradation by various proteases<sup>42</sup> that generate more porous matrices of heterogeneous rigidity. This explanation is supported by the experimental finding that matrix metalloproteinase 14-mediated local softening of the invasive front of mammary epithelial cells is necessary to overcome the stiff elastic resistance of the collagen matrix<sup>44</sup>. As tens of thousands of cancer cells differentiate on the stiff matrix, invade and then intravasate, few undifferentiated or partially differentiated cancer cells such as TRCs may follow these differentiated cancer cells and disseminate into the blood vessel. It is possible that only these soft TRCs can survive the distant organs to form metastatic colonization, the major rate-limiting step in clinically detectable macroscopic metastasis<sup>45</sup>. This interpretation may explain why metastatic

colonization is such a highly inefficient process, with only ~0.01% intravasated cancer cells being able to form metastatic colonization<sup>45</sup>. The model that soft TRCs that are much softer

than differentiated cancer cells and that are the key players in metastatic colonization is supported by a recent report that the core of human breast tumour tissues is much softer than peripheral tumour tissues and that metastatic tumour tissues in mouse lungs are much softer than surrounding normal lung tissues<sup>46</sup>. This soft cancer cell model is also supported by numerous reports over the last two decades from several labs that cancer cells, especially those metastatic cancer cells, are much softer than normal cells or non-metastatic cancer cells<sup>47–52</sup>. The long period of latency of disseminated carcinoma cells observed clinically<sup>45</sup> might be partially explained by the dormancy of the TRCs inside the uniformly stiff matrix, which restrains them from proliferating. This postulate needs to be tested in the future in animal models.

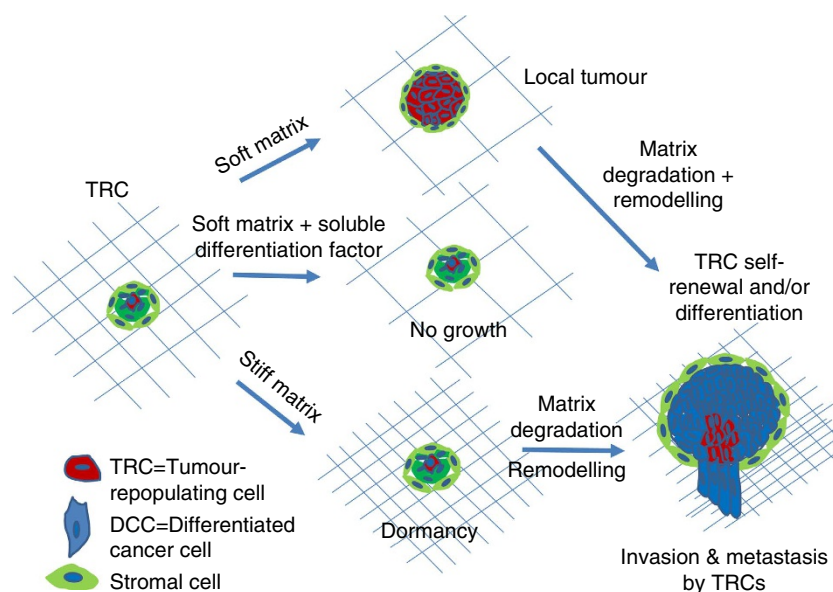


**Figure 9 | Stiff fibrin matrix elevates H3K9 methylation levels and suppresses Sox2 expression.** (a,b) Stiff fibrin matrix increases H3K9 methylation. Control cells were cultured in 90-Pa or 1,050-Pa fibrin gels for 5 days. Their H3K9 methylation level was analysed by western blotting assay (a) and quantified in b. (c) Stiff fibrin matrix suppresses Sox2 expression. Control B16-F1 cells were cultured in fibrin gels with the indicated stiffness for 5 days and then analysed for Sox2 expression by real-time RT-PCR. Mean  $\pm$  s.e.m.;  $n = 3$  independent experiments. \* $P < 0.05$ ; \*\* $P < 0.01$ .

## Methods

**Cell culture.** Murine melanoma cell line B16-F1 was purchased from American Type Culture Collection. Briefly, cells were cultured on rigid dishes with DMEM cell culture medium supplemented with 10% fetal bovine serum (Invitrogen), 2 mM L-glutamine (Invitrogen), 1 mM sodium pyruvate and 0.1 mM penicillin/streptomycin at 37 °C with 5% CO<sub>2</sub>. Cells were passaged every 3–4 days using TrypLE (Invitrogen). Glass or plastic culture dishes were coated with collagen-1 (0.2 mg ml<sup>-1</sup>) or sometimes with gelatin (0.1%) (denatured collagen) unless otherwise specified.

**3D fibrin gel preparation.** Salmon fibrinogen and thrombin were purchased from Reagent Proteins (CA, USA). Three-dimensional fibrin gels were prepared as described previously<sup>5</sup>. In brief, fibrinogen was diluted into 2 mg ml<sup>-1</sup> with T7 buffer (pH 7.4, 50 mM Tris, 150 mM NaCl). Cells were detached from 2D rigid dishes and cell density was adjusted to 10<sup>4</sup> cells per ml. Fibrinogen and cell solution mixture was made by mixing the same volume of fibrinogen solution and cell solution, resulting in 1 mg ml<sup>-1</sup> fibrinogen and 5,000 cells per ml in the mixture (for 2, 4, 8 mg ml<sup>-1</sup> (180, 420 or 1,050 Pa, respectively) fibrin gels, the initial concentration was increased to 4, 8 or 16 mg ml<sup>-1</sup>). Two hundred and fifty microlitres of cell/fibrinogen mixture was seeded into each well of 24-well plate and mixed well with pre-added 5  $\mu$ l thrombin (100 U ml<sup>-1</sup>). The cell culture plate was



**Figure 10 | A working model of melanoma tumour growth and metastasis.** TRCs proliferate in a soft matrix and eventually grow into a local tumour, while a stiff matrix restricts their proliferation and makes TRCs dormant. TRCs in the soft matrix can invade and self-renew and/or differentiate when the matrix is degraded and remodeled. When a soluble differentiation factor like retinoic acid is present, TRCs lose their proliferative property and stay quiescent even in soft matrix. However, on matrix degradation, the dormant TRCs in the original stiff matrix start to self-renew in weakened and soft sites and to differentiate in remodelled and stiffened sites, leading to invasion and eventual metastatic colonization by TRCs in very soft tissues/organs with rigidity of ~100 Pa, similar to those undifferentiated TRCs. It is known that majority of metastases from many types of solid tumours is found in very soft organs/tissues such as the lung, brain, liver and bone marrow<sup>45</sup>, but the underlying mechanism of this metastatic tropism is not understood. As the bone marrow stiffness is ~100 Pa (ref. 54), brain stiffness is ~200–1,000 Pa (ref. 54), liver stiffness is ~400 Pa (ref. 55) and lung stiffness is 100–1,000 Pa (ref. 56), all being very soft tissues, it is tempting to speculate that our model could be extended to metastases by other types of solid tumours, with implications in providing possible partial explanation to the conundrum of metastatic tropism. Clearly, this working model needs to be tested rigorously in the future.

then incubated in 37 °C cell culture incubator for 10 min. Finally, 1 ml of DMEM medium containing 10% fetal bovine serum and antibiotics was added.

**Reverse transcription-PCR and real-time RT-PCR analysis.** Total mRNA was isolated from the cells using the Trizol reagent according to the supplier's instruction (Invitrogen). Reverse transcription (RT) was performed using the TransScript First-strand cDNA Synthesis SuperMix (TransGen), according to the manufacturer's protocol. RT-PCR was performed using a PCR kit (QIAGEN), and real-time RT-PCR was performed using *GoTaq* qPCR Master Mix (Promega). The data were normalized against mouse glyceraldehyde 3-phosphate dehydrogenase. The sequences of all the primers for RT-PCR and real time RT-PCR were listed in Supplementary Tables 2 and 3.

**Chromatin immunoprecipitation assay.** We performed ChIP assay following the manufacturer's instructions (ChIP Assay Kit, Millipore). Briefly, cells were subjected to cross-linking with 1% formaldehyde in medium for 10 min at 37 °C and then lysed in SDS buffer for 10 min on ice. Chromatin was sonicated to shear DNA to an average length of 0.2–1.0 kb. Antibodies to di- and trimethyl histone H3 (Lys9) (Abcam) were used for immunoprecipitation. Normal mouse IgG (Santa Cruz) was used as negative control. The immunoprecipitation was heated to reverse the formaldehyde cross-linking and the DNA fragments in the precipitates were purified by phenol/chloroform extraction and ethanol precipitation. The immunoprecipitated DNA was quantified by real-time RT-PCR. Primers were sets corresponding to *Sox2* and glyceraldehyde 3-phosphate dehydrogenase (negative control) promoter regions. The sequences of these promoter regions were listed in Supplementary Table 4 and can be found in Transcriptional Regulatory Element Database (Cold Spring Harbor Laboratory).

**Western blotting assay.** To quantify the expression levels of G9a, SUV39h1, H3K9 di- and trimethylation, cells were lysed with 200 µl Laemmli sample buffer (Beyotime). Twenty microlitres of each sample were separated by 8–15% SDS-PAGE, blocked with 5% BSA overnight at 4 °C and incubated with primary antibodies to G9a (Rabbit, 1:1,000, Cell Signalling, 3306), SUV39h1 (Rabbit, 1:1,000, Cell Signalling, 8729), H3K9 di-methylation (Rabbit, 1:300, Millipore, 17-648) and trimethylation (Rabbit, 1:300, Millipore, 17-625) and GAPDH (Mouse, 1:1,000, Abcam, ab8245) for 2 h at room temperature. Primary antibodies were detected with goat anti-Rabbit IgG-HRP (1:2,000, Santa Cruz, sc-2004) or anti-Mouse IgG-HRP (1:2,000, Santa Cruz, sc-2005). The blots were developed using SuperSignal West Pico chemiluminescent substrate (Millipore).

**FRET probe gene construction and plasmids.** The plasmid of histone H3K9 methylation reporter (ID 22866) is purchased from Addgene (Cambridge, MA). In brief, it includes a central fragment consisting of the methyl-lysine binding domain of HP1 protein and a histone peptide, which was further fused in between an amino-terminal CFP and a carboxy-terminal YFP<sup>17</sup>. This plasmid was digested with *Bam*HI and *Sac*I, and fused to predigested pRsetB (Invitrogen) containing C-terminal YPet and NLS. The reporter containing NLS was further digested and inserted into pcDNA3 (Invitrogen) to create modified histone methylation reporter for mammalian cell expression. The insert was sequenced (W. M. Keck Center for Functional and Comparative Genomics, University of Illinois at Urbana-Champaign) to verify the integrity of the coding sequence. FRET reporter was transfected into the cells using Lipofectamine 2000 (Invitrogen) according to manufacturer's protocols.

**Magnetic twisting cytometry.** Magnetic twisting cytometry has been described elsewhere<sup>19,53</sup>. RGD-coated or poly-L-lysine-coated ferromagnetic beads (Fe<sub>3</sub>O<sub>4</sub>, 4 µm in diameter) were added to cells and then incubated for 30 min to allow for integrin binding and clustering. Beads were then magnetized in horizontal direction by applying a strong magnetic pulse (1,000 Gauss for 100 µs). A sinusoidal oscillatory waveform was applied at 0.3 Hz for 1 h to the apical surface of the cell. The amplitude of the magnetic field was varied at 35, 50 and 70 Gauss corresponding to 11, 15.9 and 22.2 Pa applied stress. The apparent applied stress is defined as the ratio of the applied torque to six times the bead volume and equals the bead constant times the applied twisting field. Cells were plated on collagen-coated (0.2 mg ml<sup>-1</sup>) or fibrinogen-coated (0.2 mg ml<sup>-1</sup>) PA gels with different rigidities or glass/plastic. Stiffness was quantified 6 h later. Cells were maintained in CO<sub>2</sub>-independent medium supplemented with 10% serum (Invitrogen) at 37 °C during experiments.

A Leica inverted microscope integrated with the magnetic twisting cytometry device and Dual-View system (Optical Insight) was used to simultaneously apply stress and capture CFP and YFP (YPet) emission images. For emission ratio imaging, the Dual-View MicroImager (Optical Insights) was used. CFP/YFP Dual EX/EM (FRET) (OI-04-SEX2) has the following filter sets: CFP: excitation, S430/25, emission S470/30; YFP: excitation, S500/20, emission S535/30. The emission filter set uses a 515-nm dichroic mirror to split the two emission images. Cells were illuminated with a 100-W Hg lamp. For FRET imaging, each CFP (1,344 pixels × 512 pixels) and each YFP image (1,344 pixels × 512 pixels) were simultaneously captured on the same screen by using a charge-coupled device

camera (C4742-95-12ERG; Hamamatsu) and a × 40, 0.55 numerical aperture air-immersion objective. A customized Matlab (Mathworks) programme was used to analyse CFP and YFP images and to obtain YFP/CFP emission ratio.

**RNA interference.** Cells were transfected with siRNA or shRNA, or complementary DNA using Lipofectamine 2000 (Invitrogen) following the manufacturer's protocol. *Silencer* Negative Control No. 1 siRNA (Invitrogen, AM4611) was used as a control in RNAi experiment. The construct sequence is 5'-GGUGAUCUUUAUGCUGUU-3' Att for G9a siRNA 1 (Invitrogen, 90322), 5'-GGUGACUUCAGAUGUGGCCctt-3' for G9a siRNA 2 (Invitrogen, 90133), 5'-GGUCCUUGUCUAUAUCAAtt-3' for SUV39h1 1 (Invitrogen, 69566), 5'-GGUGUACAACGUAUUAUCAAtt-3' for SUV39h1 2 (Invitrogen, 151927) and 5'-AGUACUGCUUACGAUACGGTt-3' for negative control siRNA. KDM7 siRNA 1 (Santa Cruz, sc-146320) is a pool of three different siRNA duplexes, sc-146320A, sc-146320B and sc-146320C, the sequence of which is 5'-GUAGUAUACCGCAGCUUAAtt-3', 5'-GUGAAUGGUAGCAUAUACAtt-3' and 5'-GAGAAUGUCUCGCCUUU CAtt-3', respectively. The sequence of KDM7 siRNA 2 is 5'-GAAUAACAUCACACUUUAAtt-3' (Invitrogen, 503916). Mif siRNA was purchased from Invitrogen (155366) and the construct sequence is 5'-GGUAUGAACACGCACUCUCtt-3'. Cdc42 siRNA was acquired from Invitrogen (catalogue number 66023) and the construct sequence is 5'-GGCAAGAGGAUUAUGACAtt-3'. *Sox2* shRNAs were obtained from Origene (TG515613). The construct sequence is 5'-GCATTACCAGAGCTAACTCAGATAGTACT-3' for scrambled shRNA, 5'-GTATAACATGATGGAGACGGAGCTGAAGC-3' for shRNA 1, 5'-AGACGCTCATGAAGAAGGAT AAGTACACG-3' for shRNA 2, 5'-AGACTACGCGACATGAACGGCTGGAGCAA-3' for shRNA 3, and 5'-AACATGATGGAGACGGAGCTGAAGCCGCC-3' for shRNA 4. The *Sox2* cDNA was obtained from Origene (MG204615). The *Cdc42*-EGFP plasmid (ID 12975) was purchased from Addgene. pEGFP-N1 vector (Clontech) was used as control.

**Traction measurement.** We followed the protocols of cell-traction quantification published previously<sup>11</sup>. In brief, cells were plated on PA gels with red fluorescent beads (0.2 µm in diameter; Molecular Probes, Invitrogen) embedded on the top surface. Before and after cell trypsinization, fluorescent images were taken to compute the displacement field of the beads using digital image correlation (DIC) in a homebuilt MATLAB program. Cell tractions were then calculated from the displacement field using inverse Boussinesq mathematical model.

**Mice experiments.** Four-week-old female C57BL/6 mice were purchased from Wuhan University Center for Animal Experiment. All animals had received humane care in compliance with the Principles of Laboratory Animal Care Formulated by the National Society of Medical Research and the guide for the National Institutes of Health of USA. The protocol was approved by the Animal Care and Use Committee of Wuhan University. B16-F1 cell spheroids were selected from 3D 90-Pa fibrin gels and pipetted to single cells. These TRCs were then transfected with *Sox2* shRNA or scrambled control via Lipofectamine 2000 (Invitrogen) following the manufacturer's protocol. These cells were harvested and the cell number was counted under microscopy. The cells were then suspended in a 0.9% NaCl solution with appropriate cell density. One hundred of TRCs transfected with *Sox2* shRNA or scrambled control were subcutaneously inoculated or intravenously injected into the tail vein of wild-type C57BL/6 mice. The tumour growth of injected mice was carefully monitored every day.

**Colony number assay.** By changing the focal planes along the z axis (the direction of gel depth), the colony number was counted view by view. At least three wells of colonies were counted per condition per day.

**Statistical analysis.** All statistics (except the Fisher's exact test for analyses of mice experiments) were performed using a two-tailed Student's *t*-test.

## References

- Visvader, J. E. & Lindeman, G. J. Cancer stem cells: current status and evolving complexities. *Cell Stem Cell* **10**, 717–728 (2012).
- Dean, M., Fojo, T. & Bates, S. Tumour stem cells and drug resistance. *Nat. Rev. Cancer* **5**, 275–284 (2005).
- Quintana, E. *et al.* Efficient tumour formation by single human melanoma cells. *Nature* **456**, 593–598 (2008).
- Quintana, E. *et al.* Phenotypic heterogeneity among tumorigenic melanoma cells from patients that is reversible and not hierarchically organized. *Cancer Cell* **18**, 510–523 (2010).
- Liu, J. *et al.* Soft fibrin gels promote selection and growth of tumorigenic cells. *Nat. Mater.* **11**, 734–741 (2012).
- Chen, J. *et al.* A restricted cell population propagates glioblastoma growth after chemotherapy. *Nature* **488**, 522–526 (2012).
- Diressens, G., Beck, B., Caauwe, A., Simons, B. D. & Blanpain, C. Defining the mode of tumour growth by clonal analysis. *Nature* **488**, 527–530 (2012).
- Schepers, A. G. *et al.* Lineage tracing reveals Lgr5+ stem cell activity in mouse intestinal adenomas. *Science* **337**, 730–735 (2012).



9. Zomer, A. *et al.* Intravital imaging of cancer stem cell plasticity in mammary tumors. *Stem Cells* **31**, 602–606 (2013).
10. Ignatius, M. S. *et al.* In vivo imaging of tumor-propagating cells, regional tumor heterogeneity, and dynamic cell movements in embryonal rhabdomyosarcoma. *Cancer Cell* **21**, 680–693 (2012).
11. Chowdhury, F. *et al.* Soft substrates promote homogeneous self-renewal of embryonic stem cells via downregulating cell-matrix tractions. *PLoS ONE* **5**, e15655 (2010).
12. Engler, A. J., Sen, S., Sweeney, H. L. & Discher, D. E. Matrix elasticity directs stem cell lineage specification. *Cell* **126**, 677–689 (2006).
13. Chen, M.-W. *et al.* H3K9 histone methyltransferase G9a promotes lung cancer invasion and metastasis by silencing the cell adhesion molecule Ep-CAM. *Cancer Res.* **70**, 7830–7840 (2010).
14. Suvà, M. L., Riggi, N. & Bernstein, B. E. Epigenetic reprogramming in cancer. *Science* **339**, 1567–1570 (2013).
15. Lakshmikuttyamma, A., Scott, S. A., DeCoteau, J. F. & Geyer, C. R. Reexpression of epigenetically silenced AML tumor suppressor genes by SUV39H1 inhibition. *Oncogene* **29**, 576–588 (2010).
16. Wagner, T. & Jung, M. New lysine methyltransferase drug targets in cancer. *Nat. Biotechnol.* **30**, 622–623 (2012).
17. Lin, C.-W., Jao, C. Y. & Ting, A. Y. Genetically encoded fluorescent reporters of histone methylation in living cells. *J. Am. Chem. Soc.* **126**, 5982–5983 (2004).
18. Lin, G. L. *et al.* Activation of beta 1 but not beta 3 integrin increases cell traction forces. *FEBS Lett.* **587**, 763–769 (2013).
19. Wang, N., Butler, J. & Ingber, D. Mechanotransduction across the cell surface and through the cytoskeleton. *Science* **260**, 1124–1127 (1993).
20. Tachibana, M. *et al.* G9a histone methyltransferase plays a dominant role in euchromatic histone H3 lysine 9 methylation and is essential for early embryogenesis. *Genes Dev.* **16**, 1779–1791 (2002).
21. Tsukada, Y., Ishitani, T. & Nakayama, K. I. KDM7 is a dual demethylase for histone H3 Lys 9 and Lys 27 and functions in brain development. *Genes Dev.* **24**, 432–437 (2010).
22. Berry, W. L. & Janknecht, R. KDM4/JMJD2 histone demethylases: epigenetic regulators in cancer cells. *Cancer Res.* **73**, 2936–2942 (2013).
23. Ballestrem, C., Hinz, B., Imhof, B. A. & Wehrle-Haller, B. Marching at the front and dragging behind: differential alphaVbeta3-integrin turnover regulates focal adhesion behavior. *J. Cell Biol.* **155**, 1319–1332 (2001).
24. Heidel, F. H. *et al.* Genetic and pharmacologic inhibition of  $\beta$ -catenin targets imatinib-resistant leukemia stem cells in CML. *Cell Stem Cell* **10**, 412–424 (2012).
25. Fong, H., Hohenstein, K. A. & Donovan, P. J. Regulation of self-renewal and pluripotency by Sox2 in human embryonic stem cells. *Stem Cells* **26**, 1931–1938 (2008).
26. Basu-Roy, U. *et al.* Sox2 maintains self renewal of tumor-initiating cells in osteosarcomas. *Oncogene* **31**, 2270–2282 (2012).
27. Adameyko, I. *et al.* Sox2 and Mitf cross-regulatory interactions consolidate progenitor and melanocyte lineages in the cranial neural crest. *Development* **139**, 397–410 (2012).
28. Kubic, J. D., Young, K. P., Plummer, R. S., Ludvik, A. E. & Lang, D. Pigmentation PAX-ways: the role of Pax3 in melanogenesis, melanocyte stem cell maintenance, and disease. *Pigment Cell Melanoma Res.* **21**, 627–645 (2008).
29. Hoashi, T. *et al.* Glycoprotein nonmetastatic melanoma protein b, a melanocytic cell marker, is a melanosome-specific and proteolytically released protein. *FASEB J.* **24**, 1616–1629 (2010).
30. Park, S. B. *et al.* SOX2 has a crucial role in the lineage determination and proliferation of mesenchymal stem cells through Dickkopf-1 and c-MYC. *Cell Death Differ.* **19**, 534–545 (2012).
31. Gangemi, R. M. R. *et al.* SOX2 silencing in glioblastoma tumor-initiating cells causes stop of proliferation and loss of tumorigenicity. *Stem Cells* **27**, 40–48 (2009).
32. Sarkar, A. & Hochedlinger, K. The sox family of transcription factors: versatile regulators of stem and progenitor cell fate. *Cell Stem Cell* **12**, 15–30 (2013).
33. Bass, A. J. *et al.* Sox2 is an amplified lineage-survival oncogene in lung and esophageal squamous cell carcinomas. *Nat. Genet.* **41**, 1238–1242 (2009).
34. Dieter, S. M. *et al.* Distinct types of tumor-initiating cells form human colon cancer tumors and metastases. *Cell Stem Cell* **9**, 357–365 (2011).
35. Esteller, M. Cancer epigenomics: DNA methylomes and histone-modification maps. *Nat. Rev. Genet.* **8**, 286–298 (2007).
36. Widschwendter, M. *et al.* Epigenetic stem cell signature in cancer. *Nat. Genet.* **39**, 157–158 (2007).
37. Guilluy, C. *et al.* Isolated nuclei adapt to force and reveal a mechanotransduction pathway in the nucleus. *Nat. Cell Biol.* **16**, 376–381 (2014).
38. Dong, C. *et al.* Interaction with Suv39H1 is critical for Snail-mediated E-cadherin repression in breast cancer. *Oncogene* **32**, 1351–1362 (2013).
39. Dong, C. *et al.* G9a interacts with Snail and is critical for Snail-mediated E-cadherin repression in human breast cancer. *J. Clin. Invest.* **122**, 1469–1486 (2012).
40. Bissell, M. J. & Hines, W. C. Why don't we get more cancer? A proposed role of the microenvironment in restraining cancer progression. *Nat. Med.* **17**, 320–329 (2011).
41. Paszek, M. J. *et al.* Tensional homeostasis and the malignant phenotype. *Cancer Cell* **8**, 241–254 (2005).
42. Levental, K. R. *et al.* Matrix crosslinking forces tumor progression by enhancing integrin signaling. *Cell* **139**, 891–906 (2009).
43. Samuel, M. S. *et al.* Actomyosin-mediated cellular tension drives increased tissue stiffness and  $\beta$ -catenin activation to induce epidermal hyperplasia and tumor growth. *Cancer Cell* **19**, 776–791 (2011).
44. Alcaraz, J. *et al.* Collective epithelial cell invasion overcomes mechanical barriers of collagenous extracellular matrix by a narrow tube-like geometry and MMP14-dependent local softening. *Integr. Biol.* **3**, 1153–1166 (2011).
45. Valastyan, S. & Weinberg, R. A. Tumor metastasis: molecular insights and evolving paradigms. *Cell* **147**, 275–292 (2011).
46. Plodinec, M. *et al.* The nanomechanical signature of breast cancer. *Nat. Nanotech.* **7**, 757–765 (2012).
47. Butler, J., Fredburg, J., Ingber, D. & Wang, N. Method and system for measurement of mechanical properties of molecules and cells. US Patent 5,486,457 (1995).
48. Guck, J. *et al.* Optical deformability as an inherent cell marker for testing malignant transformation and metastatic competence. *Biophys. J.* **88**, 3689–3698 (2005).
49. Cross, S. E., Jin, Y.-S., Rao, J. & Gimzewski, J. K. Nanomechanical analysis of cells from cancer patients. *Nat. Nanotech.* **2**, 780–783 (2007).
50. Remmerbach, T. W. *et al.* Oral cancer diagnosis by mechanical phenotyping. *Cancer Res.* **69**, 1728–1732 (2009).
51. Xu, W. *et al.* Cell stiffness is a biomarker of the metastatic potential of ovarian cancer cells. *PLoS ONE* **7**, e46609 (2012).
52. PS-OC Network. A physical sciences network characterization of non-tumorigenic and metastatic cells. *Sci. Rep.* **3**, 1449 (2013).
53. Puig-de-Morales, M. *et al.* Cytoskeletal mechanics in adherent human airway smooth muscle cells: probe specificity and scaling of protein-protein dynamics. *Am. J. Physiol. Cell Physiol.* **287**, C643–C654 (2004).
54. Discher, D. E., Mooney, D. J. & Zandstra, P. W. Growth factors, matrices, and forces combine and control stem cells. *Science* **324**, 1673–1677 (2009).
55. Georges, P. C. *et al.* Increased stiffness of the rat liver precedes matrix deposition: implications for fibrosis. *Am. J. Physiol. Gastrointest. Liver Physiol.* **293**, G1147–G1154 (2007).
56. Liu, F. & Tschumperlin, D. J. Micro-mechanical characterization of lung tissue using atomic force microscopy. *J. Vis. Exp.* **54** (2011).

## Acknowledgements

We thank Wenting Yao for help. We thank Dr Alice Ting for help with the H3K9 FRET biosensor. We thank Dr Fei Wang and his lab at UIUC for assistance. This work was supported by the funds from Huazhong University of Science and Technology and US NIH grant GM 072744 (to N.W.) and National Science Foundation through the Physics Frontiers Center Program (0822613) (to T.H.). T.H. is an investigator with the Howard Hughes Medical Institute. A.T. acknowledges partial financial support from Natural Sciences and Engineering Research Council (NSERC) of Canada through PGS Doctoral Scholarship. F.C. acknowledges support from IGB fellowship at UIUC.

## Author contributions

N.W., Y.T. and A.T. conceived the project; N.W., Y.T., A.T., J.W.C., Q.J., H.J., F.C., T.H. and B.H. designed the experiments. Y.T., A.T., J.W.C., Q.J., F.C., L.W., J.J.C., S.Z., Y.H., Y.Y., D.W., Y.Z., F.W., Y.C.P., J.S., R.S., S.D., Y.L. and L.J.L. carried out the experiments and analysed the data. N.W., Y.T., A.T., J.W.C., J.S., F.C. and Y.W. wrote the manuscript.

## Additional information

**Supplementary Information** accompanies this paper at <http://www.nature.com/naturecommunications>

**Competing financial interests:** The authors declare no competing financial interests.

**Reprints and permission** information is available online at <http://npg.nature.com/reprintsandpermissions/>

**How to cite this article:** Tan, Y. *et al.* Matrix softness regulates plasticity of tumour-repopulating cells via H3K9 demethylation and Sox2 expression. *Nat. Commun.* **5**:4619 doi: 10.1038/ncomms5619 (2014).



This work is licensed under a Creative Commons Attribution-NonCommercial-ShareAlike 4.0 International License. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in the credit line; if the material is not included under the Creative Commons license, users will need to obtain permission from the license holder to reproduce the material. To view a copy of this license, visit <http://creativecommons.org/licenses/by-nc-sa/4.0/>